Disposition and Elimination of Meropenem in Cerebrospinal Fluid of Hydrocephalic Patients with External Ventriculostomy

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The broad antibacterial spectrum and the low incidence of seizures in meropenem-treated patients qualifies meropenem for therapy of bacterial meningitis. The present study evaluates concentrations in ventricular cerebrospinal fluid (CSF) in the absence of pronounced meningeal inflammation. Patients with occlusive hydrocephalus caused by cerebrovascular diseases, who had undergone external ventriculostomy (n = 10, age range 48 to 75 years), received 2 g of meropenem intravenously over 30 min. Serum and CSF were drawn repeatedly and analyzed by liquid chromatography-mass spectroscopy. Pharmacokinetics were determined by noncompartmental analysis. Maximum concentrations in serum were 84.7 ± 23.7 μg/ml. A CSF maximum (C_{maxCSF}) of 0.63 \pm 0.50 μ g/ml (mean \pm standard deviation) was observed 4.1 \pm 2.6 h after the end of the infusion. C_{maxCSF} and the area under the curve for CSF (AUC_{CSF}) depended on the AUC for serum (AUC_S), the CSF-to-serum albumin ratio, and the CSF leukocyte count. Elimination from CSF was considerably slower than from serum (half-life at β phase $[t_{1/2\beta}]$ of 7.36 \pm 2.89 h in CSF versus $t_{1/2\beta}$ of 1.69 \pm 0.60 h in serum). The AUC_{CSF}/AUC_S ratio for meropenem, as a measure of overall CSF penetration, was 0.047 \pm 0.022. The AUC_{CSF}/AUC_{S} ratio for meropenem was similar to that for other β -lactam antibiotics with a low binding to serum proteins. The concentration maxima of meropenem in ventricular CSF observed in this study are high enough to kill fully susceptible pathogens. They may not be sufficient to kill bacteria with a reduced sensitivity to carbapenems, although clinical success has been reported for patients with meningitis caused by penicillinresistant pneumococci and Pseudomonas aeruginosa.

The carbapenems imipenem and meropenem exhibit a broad spectrum of activity against pathogens including those typically involved in bacterial central nervous system (CNS) infections. Compared to newer cephalosporins, they are active against *Listeria monocytogenes* and many anaerobes found in brain abscesses (8, 14, 29). Against listeria-induced meningitis in guinea pigs, meropenem was as effective as ampicillin plus gentamicin (15). Data for humans, however, are lacking. While imipenem-cilastatin administration has been associated with an unacceptable incidence of epileptic seizures in children with meningitis (2), meropenem does not appear to cause seizures more frequently than β -lactam antibiotics routinely used to treat meningitis (23).

In randomized clinical trials meropenem has been shown to be as effective as cefotaxime and ceftriaxone in treating community-acquired bacterial meningitis in children and adults (12, 27). For five patients with fully developed meningeal inflammation, meropenem concentrations in lumbar cerebrospinal fluid (CSF) ranged from 0.9 to 6.5 μg/ml after a single dose of 40 mg/kg of body weight. For one child with *Staphylococcus epidermidis* ventriculitis, however, the lumbar CSF meropenem concentration was only 0.3 μg/ml 6 h after dosing (5). Similarly, in the absence of meningeal inflammation lumbar CSF meropenem concentrations of 0.03 to 0.26 μg/ml were observed 0.5 to 2.5 h after a dose of 1 g (11).

In the early stages of meningococcal meningitis and in *Streptococcus pneumoniae* sepsis after splenectomy, ventricular shunt

infections, and corticoid-treated meningitis, bacteria are present in the subarachnoid space in the absence of pronounced inflammation (3, 9, 10, 13, 22, 24). Furthermore, ventricular antimicrobial concentrations may be up to 10 times lower than lumbar levels after intravenous (i.v.) administration (1). For these reasons, successful treatment of bacterial CNS infections must ensure sufficient drug concentrations in lumbar and ventricular CSF under conditions of not only severe but also minor impairment of the blood-CSF barrier. In the present study we investigated the pharmacokinetics of meropenem in the ventricular CSF of patients with external ventriculostomy and minor abnormalities of the blood-CSF barrier.

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MATERIALS AND METHODS

Ten patients (five females and five males; age range, 48 to 75 years; serum creatinine concentration, 0.7 to 1.7 mg/dl) suffering from extracerebral infections caused by bacteria of proven or presumed (when treatment had to be initiated before susceptibility data were available) susceptibility to meropenem were included in this study. They had undergone external ventriculostomy due to occlusive hydrocephalus caused by cerebrovascular accidents. Patients with clinical evidence of ventriculitis and with serum creatinine concentrations above 2 mg/dl were not considered.

The CSF leukocyte count on the day of meropenem administration ranged from 0 to 91 cells/µl (median, 8 cells/µl), and the CSF protein content ranged from 80 to 1,716 µg/ml (median, 587 µg/ml). All patients received a first dose of 2 g of meropenem i.v. (Meronem; Zeneca GmbH, Plankstadt, Germany) over a 30-min period. Treatment was continued 16 h later with 1 g of meropenem three times a day. For further information on the patients studied see Table 1.

Ventricular CSF and arterial blood were sampled from indwelling catheters before, at the end, and at 30 min and 1, 2, 4, 7, 10, 13, and 16 h after the end of the infusion. At 10 min after the end of the infusion, only blood was drawn. The

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TABLE 1	Characteristics	of the	patients investigated	а

	Ago			Т:		Serum		Serum	CSF analysis ^d			
Pa- tient	Age (yr), sex	r), wt Disease interval Treatment		creatinine concn (mg/dl)	CL _{CR} ^c (ml/min)	albumin concn (g/liter)	Protein concn (mg/liter)	$\begin{array}{c}Q_{\rm Alb}\\(10^{-3})\end{array}$	WBC/ mm ³	RBC/ mm ³		
1	48, m	85	Intracerebral hemorrhage	4	Amoxicillin, clavulanate, genta- micin, metronidazole	1.7	64	28	425	5.4	91	8,875
2	75, f	70	Subarachnoid hemorrhage	12		0.7	76	23	284	5.3	5	20
3	63, m	80	Intracerebral hemorrhage	4	Mannitol	0.7	122	27	1,630	29.9	68	2,560
4	75, f	75	Cerebellar hemorrhage	7	Mannitol	0.8	72	28	848	10.1	5	1,605
5	55, f	85	Subarachnoid hemorrhage	8	Dexamethasone, mannitol	0.7	121	27	518	3.0	26	4,096
6	52, m	70	Basilar artery occlusion	3	Mannitol, amoxicillin, genta- icin, metronidazole	0.8	107	26	201	4.4	1	40
7	63, f	80	Infratentorial infarction	5	Mannitol, amoxicillin, clavu- lanate	0.9	81	29	80	1.5	0	20
8	66, m	95	Intracerebral hemorrhage	11	Mannitol, cefotaxime, metro- nidazole	0.8	122	31	1,716	4.9	11	4,690
9	55, m	95	Intracerebral hemorrhage	8	Dexamethasone	0.7	160	29	840	7.8	44	7,851
10	60, f	65	Intracerebral hemorrhage	8	Glycerol, cefotaxime	0.9	68	22	843	6.1	3	1,840

^a Patient order reflects descending peak CSF meropenem concentration.

^d WBC, leukocyte; RBC, erythrocyte.

ventricular CSF samples were drawn from the port nearest to the site of insertion of the ventriculostomy, and 1 ml of CSF was discarded before the CSF sample (1 ml) was collected. Blood was allowed to clot for 3 min. Then blood and CSF were centrifuged immediately. The supernatants were frozen at -70° C.

Informed consent to participation in this study was obtained from the nearest relative. The study protocol was approved by the Ethics Committee of the Medical Faculty of the University of Göttingen, Göttingen, Germany.

After deproteinization with acetonitrile, serum and CSF drug concentrations were measured by liquid chromatography-mass spectroscopy/mass spectroscopy (LC-MS/MS). A total of 50 μ l of each sample was separated on a reversed-phase column (40 by 4.6 mm) with an isocratic system. The mobile phase consisted of 78% 0.005 M ammonium acetate buffer and 22% acetonitrile. Peaks were monitored by a selected reaction monitoring method as follows: precursor (meropenem + H)⁺ \rightarrow product ion for meropenem m/z 384 \rightarrow m/z 68, and for the internal standard, m/z 518 \rightarrow m/z 143. Meropenem eluted after 0.9 min, and the internal standard eluted after 1.5 min. Mac Quan software (version 1.4-noFPU, 1991 to 1995) (Perkin-Elmer, Toronto, Canada) was used for the evaluation of the chromatograms.

The serum and CSF samples were measured against a calibration row prepared from drug-free serum and CSF, respectively. No interferences in serum and CSF were observed for meropenem or the internal standards. Calibration was performed by weighted (1/concentration) linear regression. The linearity of the meropenem calibration curve for serum was demonstrated between 0.019 and 42.1 μ g/ml. For CSF, linearity was found in a concentration range of 0.0042 to 4.91 μ g/ml. The lowest calibration levels were taken as quantification limits. Samples with drug concentrations above the quantification limits were prediluted with tested drug-free plasma.

For control of interassay variation, spiked quality control samples were prepared by adding defined amounts of meropenem to tested drug-free serum and CSF. The interassay precisions of the spiked meropenem quality controls in serum were 4.1% at a drug concentration of 30.1 μ g/ml, 3.3% at 3.01 μ g/ml, 5.4% at 0.343 μ g/ml, and 4.9% at 0.041 μ g/ml. The interassay precisions of the spiked quality controls in CSF were 1.1% at a concentration of 3.94 μ g/ml, 2.3% at 0.541 μ g/l, 3.7% at 0.075 μ g/ml, and 3.1% at 0.012 μ g/ml. The accuracy of the meropenem standards ranged from 95.1 to 104.3% for serum and from 97.5 and 10.4.3% for CSF.

Concentration-time curves for serum and CSF were analyzed by noncompartmental pharmacokinetic methods using the program Topfit 2.0 (Gödecke-Schering-Thomae, Biberach an der Riss, Germany). Peak serum and CSF drug concentrations ($C_{\rm maxS}$ and $C_{\rm maxCSF}$, respectively), time from the end of the infusion to the respective peak concentrations ($t_{\rm maxS}$ and $t_{\rm maxCSF}$), and time above hypothetical MICs of 0.125 and 0.5 µg/ml in CSF were directly taken from the concentration-time curves. Elimination rate constants ($k_{\rm el}$) were determined by log-linear regression analysis [weighting function $g(y_i)=1/y_i$, where $g(y_i)$ is the weighted concentration and y_i is the individual concentration as determined by LC-MS/MS], and half-lives at β phase ($t_{1/2\beta}$) were determined as $\ln 2/k_{\rm el}$. The areas under the concentration-time curves for serum and CSF up to the last measurable drug concentration ($\Delta UC_{\rm SD_4}$, and $\Delta UC_{\rm CSF_{\rm el}}$, respectively) were estimated by the linear trapezoidal rule. Extrapolation to infinity ($\Delta UC_{\rm S}$, and the ratio $\Delta UC_{\rm CSF}$ was expressed as a percentage. The total clearance

in serum (CL) was calculated as dose/AUC_S, and the apparent volume of distribution ($V_{\rm B}$) was calculated as dose/(AUC_S · $k_{\rm cl}$). The volume of distribution in steady state ($V_{\rm SS}$) was determined as CL · [mean residence time (infusion time/2)], where the mean residence time is defined as $\int_0^\infty C_{\rm S}(t)$ · tdt/AUC_S. The exit rate constant from CSF ($k_{\rm outCSF}$ or CL_{outCSF}/ $V_{\rm CSF}$) was determined by plotting $\Delta C_{\rm CSF}$ versus AUC_{CSF}/AUC_S · AUC_{Stn-tip+1} - AUC_{CSF,n-tip+1}. Linear least-square regression yielded a straight line without intercept, the slope equalling $k_{\rm outCSF}$ (26). $k_{\rm outCSF}$ can be used to estimate the approximate elimination half-life from CSF after intraventricular injection of meropenem ($t^*_{1/2CSF}$) by using the equation $k_{\rm outCSF} = \ln 2/t^*_{1/2CSF}$.

Pharmacokinetic data were expressed as means \pm standard deviations (SD). Medians are provided to facilitate comparison with previous data on CSF penetration by other compounds, where normal distributions were not always present. The determinants of the entry of meropenem into CSF were analyzed by stepwise multiple linear regression with AUC_{CSF} and C_{maxCSF} as dependent variables and AUC_S, C_{maxS}, V_{SS} , $t_{1/2BS}$, age, serum creatinine, CSF protein content, CSF-to-serum albumin ratio (Q_{Alb}), time interval between insertion of external ventriculostomy and the first meropenem infusion, and CSF leukocyte count as independent variables.

RESULTS

At the end of the infusion, maximum serum meropenem concentrations were 84.7 \pm 23.7 µg/ml (mean \pm SD). CL was 270 \pm 137 ml/min, and the $V_{\rm SS}$ amounted to 28.7 \pm 6.8 liters (Table 2). The mean maximum CSF drug concentration of 0.63 \pm 0.50 µg/ml was observed 4.1 \pm 2.6 h after the end of the infusion. The lowest individual patient CSF maximum observed was 0.13 µg/ml, and the highest was 1.60 µg/ml. Elimination from CSF was considerably slower than from serum (t_{1/2B} of 7.36 \pm 2.89 h in CSF versus $t_{1/2B}$ of 1.69 \pm 0.60 h in serum) (Fig. 1). The AUC $_{\rm CSF}/{\rm AUC}_{\rm S}$ ratio for meropenem, as a measure of overall CSF penetration, was 0.047 \pm 0.022 (Table 3). $k_{\rm outCSF}$ ranged from 0.056 to 0.321/h, corresponding to an approximate elimination half-life from CSF, which would be effective after intraventricular injection of meropenem ($t^*_{\rm 1/2CSF}$), of 6.75 \pm 3.44 h (Table 3).

In the case of highly susceptible bacteria requiring a MIC of 0.125 μ g/ml, the time above the MIC in CSF would be 12 \pm 5 h. In the presence of a moderately susceptible bacterium requiring a MIC of 0.5 μ g/ml the CSF meropenem concentrations would be below the MIC for the whole period of observation for six patients. For the other four patients the time above the MIC would range from 9 to 13 h (Table 3).

Stepwise multiple linear regression with AUC_{CSF} as the de-

^b Time interval between insertion of external ventriculostomy and the first meropenem infusion.

^c Creatinine clearance (CL_{CR}) was estimated by the following equations: males, (140 – age) · weight · 72⁻¹ · serum creatinine⁻¹; females, (140 – age) · weight · 85⁻¹ · serum creatinine⁻¹. These equations tend to overestimate creatinine clearance in obese and emaciated patients.

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$C_{\rm max}$ (mg/l)	AUC (mg · h/liter)	CL (ml/min)	CL/kg (ml/min/kg)	$t_{1/2\beta}$ (h)	V_{β} (liter)	$V_{\rm SS}$ (liter)	V _{SS} /kg
90.6	210.6	158	1.86	2.72	37.2	31.9	0.38
32.4	305.1	109	1.56	2.34	22.2	19.8	0.28
96.1	206.4	162	2.03	1.11	15.5	28.1	0.35

TABLE 2. Kinetic properties of meropenem in serum after i.v. administration of 2 g

$C_{\max} \pmod{l}$	AUC (mg · h/liter)	CL (ml/min)	CL/kg (ml/min/kg)	$t_{1/2\beta}$ (h)	V_{β} (liter)	$V_{\rm SS}$ (liter)	V _{SS} /kg
90.6	210.6	158	1.86	2.72	37.2	31.9	0.38
132.4	305.1	109	1.56	2.34	22.2	19.8	0.28
96.1	206.4	162	2.03	1.11	15.5	28.1	0.35
80.5	234.3	142	1.89	2.32	28.6	26.4	0.35
82.1	91.2	365	4.29	1.32	41.8	24.6	0.29
90.6	135.1	247	3.53	1.24	26.5	25.2	0.36
56.9	130.0	256	3.20	1.96	43.5	34.5	0.43
101.6	97.1	343	3.61	1.01	30.0	19.9	0.21
65.6	60.7	549	5.78	1.44	68.7	37.8	0.40
51.0	90.2	369	5.68	1.41	45.2	38.9	0.60
84.7 (23.7)	156.1 (78.9)	270 (137)	3.34 (1.55)	1.69 (0.60)	35.9 (15.0)	28.7 (6.8)	0.37 (0.10)
86.4	132.6	252 `	3.37	1.43	33.6	27.3	0.36
	(mg/l) 90.6 132.4 96.1 80.5 82.1 90.6 56.9 101.6 65.6 51.0 84.7 (23.7)	(mg/l) (mg · h/liter) 90.6 210.6 132.4 305.1 96.1 206.4 80.5 234.3 82.1 91.2 90.6 135.1 56.9 130.0 101.6 97.1 65.6 60.7 51.0 90.2 84.7 (23.7) 156.1 (78.9)	(mg/l) (mg · h/liter) (ml/min) 90.6 210.6 158 132.4 305.1 109 96.1 206.4 162 80.5 234.3 142 82.1 91.2 365 90.6 135.1 247 56.9 130.0 256 101.6 97.1 343 65.6 60.7 549 51.0 90.2 369 84.7 (23.7) 156.1 (78.9) 270 (137)	(mg/l) (mg · h/liter) (ml/min) (ml/min/kg) 90.6 210.6 158 1.86 132.4 305.1 109 1.56 96.1 206.4 162 2.03 80.5 234.3 142 1.89 82.1 91.2 365 4.29 90.6 135.1 247 3.53 56.9 130.0 256 3.20 101.6 97.1 343 3.61 65.6 60.7 549 5.78 51.0 90.2 369 5.68 84.7 (23.7) 156.1 (78.9) 270 (137) 3.34 (1.55)	(mg/l) (mg · h/liter) (ml/min) (ml/min/kg) I _{1/2β} (h) 90.6 210.6 158 1.86 2.72 132.4 305.1 109 1.56 2.34 96.1 206.4 162 2.03 1.11 80.5 234.3 142 1.89 2.32 82.1 91.2 365 4.29 1.32 90.6 135.1 247 3.53 1.24 56.9 130.0 256 3.20 1.96 101.6 97.1 343 3.61 1.01 65.6 60.7 549 5.78 1.44 51.0 90.2 369 5.68 1.41 84.7 (23.7) 156.1 (78.9) 270 (137) 3.34 (1.55) 1.69 (0.60)	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$

^a SD, standard deviation.

pendent variable yielded three variables, which explained 98% of the variance (P < 0.00005): AUC_S, Q_{Alb} , and CSF leukocyte density. Similarly, for the dependent variable C_{maxCSF}, AUC_s, CSF leukocyte count, and Q_{Alb} explained 97% of the variance (P < 0.00005) (Table 4). The inclusion of additional independent variables (V_{SS} , $t_{1/2BS}$, age, serum creatinine, CSF protein content, time interval between insertion of external ventriculostomy and the first meropenem infusion) only led to small, nonsignificant increases of the goodness of fit (R^2) .

DISCUSSION

Meropenem is a weak acid with a pK_a of 2.9. At physiological pH, it is highly ionized and hydrophilic (manufacturer's information). The molecular mass of the anhydrous free acid is 383.5 Da and that of the trihydrate is 437.5 Da. Binding to serum proteins is neglegibly low (11) and should not affect CSF penetration (21). As with other hydrophilic compounds with similar molecular masses (16, 18, 19), maximum concentrations in CSF were observed several hours after the end of the meropenem infusion and drug elimination was substantially slower from CSF than from serum. In the present study, after a single dose of 2 g of meropenem, maximum concentrations in CSF (median, 0.45 µg/ml) were almost identical to those observed after a 2-g i.v. dose of cefotaxime or ceftriaxone (median, 0.44 and 0.43 µg/ml, respectively [19]). Meropenem CSF maxima were lower than maximum CSF concentrations encountered after the administration of 3 g of ceftazidime (18) or 6 g of piperacillin (16). The AUC_{CSF}/AUC_S ratio for meropenem (0.046) compared well with the median AUC_{CSF}/AUC_S ratios for ceftazidime (0.054) (18) and piperacillin (0.034) (16) but was two- to threefold below that for cefotaxime (0.12) and approximately seven times above the median AUC_{CSF}/AUC_S ratio for ceftriaxone (0.007) (19). We are unaware of a comparable study concerning the penetration of imipenem into the ventricular CSF. An investigation of imipenem in lumbar CSF 1, 2, 4, 6, and 8 h after i.v. administration of 1 g of imipenem and 1 g of cilastatin yielded mean imipenem concentrations of 0.62 to 0.90 µg/ml, i.e., concentrations similar to those found by us in ventricular CSF after 2 g of meropenem. The $AUC_{CSF_{1-8h}}/AUC_{S_{1-8h}}$ ratio for imipenem in the absence of meningeal inflammation was 0.073. Extrapolation to infinity was not performed. In the presence of meningeal inflammation, lumbar CSF imipenem concentrations were two to three times higher than those without inflamed meninges (6).

In all patients, elimination from the CSF space was slower than elimination from serum. To assess the effect of the

passage of drug from blood to CSF during the elimination phase on $t_{1/2BCSF}$, the exit rate constant k_{outCSF} was calculated allowing an estimate of the elimination half-life from CSF, which would be effective after intraventricular drug injection $(t^*_{1/2\text{CSF}})$. Independent of $t_{1/2\beta S}$, the $t^*_{1/2\text{CSF}}$ s should be almost identical for hydrophilic drugs predominantly cleared from CSF by bulk flow. On average, $t^*_{1/2\text{CSF}}$ was 0.6 h shorter than $t_{1/2\beta {\rm CSF}}$ (Table 3). The relatively long elimination half-life of meropenem in CSF suggests accumulation after repeated dosing. Assuming linear kinetics, mean CSF drug concentrations in steady state can be estimated from the AUC_{CSF} after the first dose by dividing AUC_{CSF} by the dosing interval (17). The mean AUC_{CSF} of meropenem was 8.55 mg · h/liter. Repeated doses of 2 g and dosing intervals of 8 h, i.e., the maximum daily dose recommended by the manufacturer, would result in mean steady state concentrations in CSF of approximately 1.1 µg/ml.

In the present study, stepwise multiple linear regression analysis using both AUC_{CSF} and C_{maxCSF} as dependent variables demonstrated the impact of AUC_S upon penetration into CSF. Consistently, the patient with an elevated serum creati-

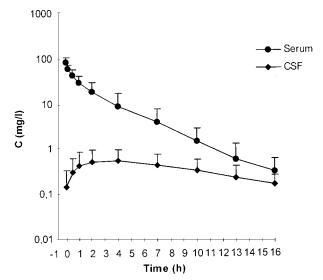


FIG. 1. Concentration-time curves of meropenem in serum and CSF after i.v. administration of 2 g in patients with external ventriculostomy. Note the lag time between maximum serum and CSF meropenem concentrations and the slow drug elimination from CSF. Data are means (± SD) for 10 patients.

TABLE 3. Kinetic properties of meropenem is	n CSF after i	i.v. administration of 2 g
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Patient	$C_{ m max}$ (mg/liter)	T _{max} ^a (h)	AUC (mg · h/liter)	AUC _{CSF extrapolated} / AUC _{CSF} (%)	t _{1/2β} (h)	AUC _{CSF} / AUC _S	Time (h) above a MIC of:		t* _{1/2CSF} (h)	
	(mg/iter)	(11)	(mg m/mer)	AUC _{CSF} (%)		AUC_S	0.125 μg/ml	0.5 μg/ml	(11)	
1	1.60	2	14.2	16.1	5.65	0.067	16	9	2.18	
2	1.22	4	17.2	33.1	8.55	0.056	16	11	5.68	
3	1.09	4	18.3	30.3	7.79	0.089	16	13	8.46	
4	0.65	7	14.7	43.7	11.9	0.063	16	12	10.66	
5	0.48	4	4.46	9.8	4.57	0.049	15	0	4.63	
6	0.41	10	5.80	14.7	6.46	0.043	13	0	8.61	
7	0.29	2	4.87	30.6	8.31	0.037	14	0	7.87	
8	0.24	2	2.27	14.1	5.30	0.023	8	0	4.78	
9	0.20	2	1.18	4.2	3.31	0.019	3	0	2.16	
10	0.13	4	2.54	44.4	11.8	0.028	3	0	12.47	
Mean (SD ^b) Median	0.63 (0.50) 0.45	4.1 (2.6) 4	8.55 (6.73) 5.34	26.1 (16.4) 23.4	7.36 (2.89) 7.13	0.047 (0.022) 0.046	12 (5) 14	5 (6) 0	6.75 (3.44) 6.77	

 $^{^{}a}_{t}T_{\text{max}}$, time to maximum concentration.

nine concentration and the two oldest patients ranked first, second, and fourth with respect to the maximum CSF concentration and second, third, and fourth with respect to $\mathrm{AUC}_{\mathrm{CSF}}.$ Parameters describing the state of the blood-CSF barrier (Q_{Alb} and CSF leukocyte count) also had a substantial influence on CSF penetration. In the case of $\mathrm{AUC}_{\mathrm{CSF}},$ the Q_{Alb} was more significant than the CSF leukocyte count in explaining the interindividual variance of the entry of meropenem into CSF. The patient with the greatest Q_{Alb} ranked first with respect to $\mathrm{AUC}_{\mathrm{CSF}}.$ With $\mathrm{C}_{\mathrm{maxCSF}},$ the increase of R^2 was greater with the CSF leukocyte count than with the $Q_{\mathrm{Alb}}.$ In both cases, the increase of R^2 achieved by the inclusion of the third independent variable was small but significant.

In a previous study of lumbar CSF during severe meningeal inflammation, meropenem concentrations after a single dose of 40 mg/kg (0.9 to 6.5 μ g/ml) were higher than those found by us (5). The two probable reasons are as follows: (i) disruption of the blood-CSF barrier of our patients, as estimated by the CSF protein content and the Q_{Alb} , ranged from absent to moderate and (ii) in the present study, meropenem concentrations were determined in ventricular CSF. The current practice of measuring lumbar CSF antibiotic concentrations and assuming identical antibiotic concentrations in all parts of the CSF compartment often overestimates the antibiotic concentrations in the CSF surrounding the brain. In a recent study of primates the AUC of lamivudine was approximately five times higher in the lumbar than in the ventricular CSF after i.v. infusion (1). For these reasons, the CSF drug concentrations measured by us are probably the minimum concentrations which can be encountered in bacterial CNS infections.

The maximum CSF meropenem concentrations of all patients included in this study equalled or surmounted the MICs at which 90% of isolates are inhibited (MIC₉₀s) for *Neisseria meningitidis*, *Haemophilus influenzae*, penicillin-sensitive *Streptococcus pneumoniae*, and for most members of the family *Enterobacteriaceae* (\leq 0.125 µg/ml) (8, 29). For the majority of patients, the CSF maxima were above the MIC₉₀s of *L. monocytogenes*, methicillin-sensitive *Staphylococcus aureus*, and *Serratia marcescens* strains (0.25 µg/ml). Less than half of our patients, however, reached a CSF maximum corresponding to the MIC₉₀s of *Bacteroides* spp. (0.5 µg/ml) and penicillin-resistant *S. pneumoniae* (0.5 and 1 µg/ml) (8, 25, 29). Nevertheless, three cases of *S. pneumoniae* meningitis caused by strains for which the MICs were 0.25 to 1 µg/ml were successfully treated

with meropenem (30). The MIC $_{90}$ of *P. aeruginosa* and *Enterococcus* spp. (8 µg/ml) (8, 29) was considerably higher than the maximum CSF concentration encountered in this study. Again, two cases of successful treatment of *Pseudomonas* meningitis with meropenem have been documented (4, 7). The MIC $_{90}$ of meropenem for *Staphylococcus epidermidis* was 4.3 µg/ml (29). Methicillin resistance frequently encountered in *S. epidermidis* precludes the use of any β -lactam antibiotic. Therefore, meropenem is only suitable for methicillin-sensitive strains with documented susceptibility.

In the case of a fully susceptible bacterium for which the meropenem MIC is 0.125 µg/ml, the meropenem concentration in CSF would be above the MIC for most of the observation period (12 ± 5 h). However, for 6 of 10 patients concentrations in CSF did not reach 0.5 µg/ml (Table 3). Obviously, with the CSF levels observed, a meropenem concentration/ MIC ratio above 10, which is probably required for a rapid bactericidal effect in experimental meningitis (20, 28), will only be reached in the presence of highly susceptible pathogens. Yet, the minimum CSF antibiotic concentration in relation to the MIC and MBC essential for successful eradication of bacteria remains to be determined. An exact MIC determination is mandatory for CNS infections in order to assess whether sufficient CSF drug concentrations can be counted upon. In selected cases the measurement of CSF drug concentrations may be necessary. To overcome the problem of minor meningeal inflammation in bacterial CNS infections, more lipophilic compounds, whose CSF penetration is less dependent on the functional state of the blood-CSF barrier, may be used in combination with cephalosporins or carbapenems. This strat-

TABLE 4. Stepwise multiple linear regression analysis of the entry of meropenem into ventricular CSF

Dependent variable	Independent variable	R^2	F value	P value
AUC _{CSF}	AUCs	0.87	53.4	0.0001
	$Q_{ m Alb}$	0.97	100.3	< 0.00005
	CSF leukocyte density	0.98	131.0	< 0.00005
C_{maxCSF}	AUC_S	0.64	14.2	0.0055
marcor	CSF leukocyte density	0.94	59.2	< 0.00005
	$Q_{ m Alb}$	0.97	69.5	< 0.00005

^b SD, standard deviation.

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egy has been advocated for dexamethasone-treated patients with bacterial meningitis (24).

In conclusion, in the presence of minor impairment of the blood-CSF barrier, the CSF maxima of meropenem in ventricular CSF were high enough to kill fully susceptible pathogens. They may not be sufficient for bacteria with a reduced sensitivity to carbapenems, although clinical success has been reported for patients with meningitis caused by penicillin-resistant pneumococci and P. aeruginosa. From a pharmacokinetic view, increasing the dose of meropenem to 8 to 12 g/day, as is current practice with other β-lactam antibiotics, would be desirable in these conditions. We are, however, unaware of data which would indicate that such high doses of meropenem would be tolerated without toxic side effects. The AUC_{CSF}/ AUC_s ratio for meropenem, as a measure of overall CSF penetration, was similar to that for β -lactam antibiotics with a low binding to serum proteins. The elimination half-life in CSF of approximately 7 h suggests moderate accumulation after repeated dosing.

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